

Tel 24hrs: +44 (0) 24 - 76 - 323222 Fax / Ans: +44 (0) 24 - 76 - 323333

University of Warwick Science Park, Venture Centre, Sir William Lyons Road, Coventry CV4 7EZ Website: www.micropathology.com E-mail: info@micropathology.com

M. genitalium resistance assays (macrolide and fluoroquinolone)

Mycoplasma genitalium (commonly referred to as MGen and also known as *Mycoplasmoides genitalium*^{1,2})* is a bacterium emerging as a significant causative agent of sexually transmitted infections in both males and females³. *M. genitalium* was initially isolated from the urethral swabs of males with non-gonococcal urethritis in 1981 and was identified as a new species in 1983⁴. Infection is closely associated with urethritis in men, and cervicitis in women, with links also been made to pelvic inflammatory disease. Through use of polymerase chain reaction (PCR), *M. genitalium* has been established as a key source of non-chlamydial, non-gonococcal urethritis in men and mucopurulent cervicitis in women⁵. The bacterium is one of the smallest known self-replicating organisms that proliferates on the skin cells of the human genital and urinary tracts. Bacterial infections such as those caused by *M. genitalium* are treated with antibiotics, however recent studies have found alarming rates of resistance to macrolide treatments. Recent changes to the British Association for Sexual Health and HIV (BASHH) guidelines have stated that all patient samples containing *M. genitalium* DNA should, where feasible, additionally be tested for macrolide resistance mediating mutations. Where macrolide resistance associated mutations exist treatment is typically followed up with a fluoroquinolone antimicrobial.

Current recommendations suggest treatment for uncomplicated infections with Doxycycline 100 mg bd for seven days followed by azithromycin (macrolide) 1g orally as a single dose then 500 mg orally once daily for 2 days where organism is known to be macrolide-sensitive or where resistance status is unknown. Where macrolide resistance is identified, in azithromycin failure or complicated infections treatment should be with Moxifloxacin (fluoroquinolone) 400 mg orally once daily for 7 days or 14 days respectively⁶. Treatment failure of this second-line option has also been reported and is likely to increase⁷. Recent studies suggesting that moxifloxacin success rates range from 69 to 100%⁸. Therefore, detection of resistance mutations may become increasingly important in the treatment of these infections.

Signs and Symptoms

M. genitalium infection yields a vast range of clinical symptoms, though asymptomatic colonisation may occur. Infection can cause urethral inflammation, referred to as urethritis, in both men and women, which can result in pain when urinating (dysuria), the frequent need to urinate and pain during sex. Resistance should be suspected when the symptoms or the organism persists despite appropriate therapy.

Samples and testing pathway

Micropathology Ltd. accept both genital swabs and urine (males only) as UKAS accredited sample types.

Where macrolide resistance testing is requested, all samples will be routinely tested for *M. genitalium* DNA in the first instance.

Macrolide resistance assay (Mg23)

Macrolides are bacteriostatic antibiotics that by binding to the 50S subunit of the ribosome and inhibit bacterial protein synthesis of a range of bacteria including Grampositives. Mutations associated with macrolide resistance in *M. genitalium* have been identified at positions 2058 and 2059 (*Escherichia coli* numbering) on region V of the 23S rRNA gene. At Micropathology Ltd., our in-house single round assay amplifies a region of the 23S rRNA gene and uses sequencing to interrogate the bases found at positions 2058 and 2059. Where changes associated with macrolide resistance are observed, results are reported as "Resistance associated mutation(s) observed", followed by the mutation.

Fluoroquinolone resistance assay (ParC)

Fluoroquinolones target topoisomerase enzymes responsible for regulation of DNA supercoiling. Mutations in the quinolone resistance determining region (QRDR) of the *parC* gene (encoding the ParC subunit A of topoisomerase IV) have been associated with resistance to fourth generation fluoroquinolones such as moxifloxacin. The best characterised resistance associated mutations cause changes in serine 83 (S83), and aspartic acid 87 (D87) (*M. genitalium* numbering) in the QRDR of the *parC* gene. Other single nucleotide polymorphisms (SNPs) in the *parC* and *gyrA* (encodes subunit A in DNA gyrase) may also contribute to fluoroquinolone treatment failure, however their role in mediating resistance is currently not well defined⁷.

At Micropathology Ltd. our single round ParC PCR assay amplifies a region of *parC* in *M. genitalium*. Analysis is based upon interrogation of the sequence data of the *M. genitalium parC* gene only, which confer amino acid changes in S83 and D87 associated with resistance to fluoroquinolones. Not all mutations capable of causing fluoroquinolones resistance are known and/or are fully characterised. Consequently, inferred resistance based upon genotypic data may not correlate directly with empirically determined resistance based upon phenotypic analyses.

Where changes associated with fluoroquinolone resistance are observed, results are reported as "Resistance associated mutation(s) observed", followed by the mutation.

*At this time the microbiology/clinical microbiology community has not agreed or defined whether the use of *Mycoplasma* or *Mycoplasmoides* is the most appropriate nomenclature to define the similarities and differences within the former *Mycoplasma* genus. To reduce clinical confusion, we will refer to the organism as its original designation of *Mycoplasma genitalium* but acknowledge that either classification conforms with current practices.

References

¹Munson, E and Carroll, K.C. 2021. Summary of Novel Bacterial Isolates Derived from Human Clinical Specimens and Nomenclature Revisions Published in 2018 and 2019. *Clinical Microbiology*, 59:2

² Mitchell Balish, Assunta Bertaccini, Alain Blanchard, Daniel Brown, Glenn Browning, Victoria Chalker, Joachim Frey, Gail Gasparich, Ludwig Hoelzle, Tom Knight Jr, Christine Knox, Chih-Horng Kuo, Lucia Manso-Silván, Meghan May, J. Dennis Pollack, Ana S. Ramírez, Joachim Spergser, David Taylor-Robinson, Dmitriy Volokhov, Yan Zhao. 2019. Recommended rejection of the names Malacoplasma gen. nov., Mesomycoplasma gen. nov., Metamycoplasma gen. nov., Metamycoplasma gen. nov., Mycoplasmoidales ord. nov., Mycoplasmoides gen. nov., Mycoplasmoidales ord. nov., Mycoplasmoides gen. nov., International journal of systematic and evolutionary microbiology. 69:11

³ Sethi, S., Singh, G., Samanta, P. and Sharma, M. 2012. *Mycoplasma genitalium*: An emerging sexually transmitted pathogen. *Indian Journal of Medical Research*, vol. 136, no. 6, pp. 942-955.

⁴ Daley, G., Russell, D., Tabrizi, S. and McBride, J. 2014. *Mycoplasma genitalium*: a review. *International Journal of STD and AIDS*, vol. 25, no. 7.

⁵ Falk, L., Fredlund, H. and Jensen, JS. 2005. Signs and symptoms of urethritis and cervicitis among women with or without *Mycoplasma genitalium* or *Chlamydia trachomatis* infection. *Sexually Transmitted Infections*, vol. 81, no. 1, pp. 73-78.

⁶ Soni, S., Horner, P., Rayment, M., Pinto-Sander, N., Naous, N., Parkhouse, A., Bancroft, D., Patterson, C. and Fifer, H., 2019. British Association for Sexual Health and HIV national guideline for the management of infection with Mycoplasma genitalium (2018). *International Journal of STD & AIDS*, 30(10), pp.938-950.

7Couldwell, D., Tagg, K., Jeoffreys, N. and Gilbert, G., 2013. Failure of moxifloxacin treatment in *Mycoplasma genitalium* infections due to macrolide and fluoroquinolone resistance. *International Journal of STD & AIDS*, [online] 24(10), pp.822-828.

⁸Manhart, L., Skov Jensen, J., Bradshaw, C., Golden, M. and Martin, D., 2015. Efficacy of antimicrobial therapy for Mycoplasma genitalium infections. Clinical Infectious Diseases, 61(8), pp.S802–S817.

⁹Hamasuna, R., Le, P., Kutsuna, S., Furubayashi, K., Matsumoto, M., Ohmagari, N., Fujimoto, N., Matsumoto, T. and Jensen, J., 2018. Mutations in ParC and GyrA of moxifloxacin-resistant and susceptible *Mycoplasma genitalium* strains. *PLOS ONE*, [online] 13(6), p.e0198355.